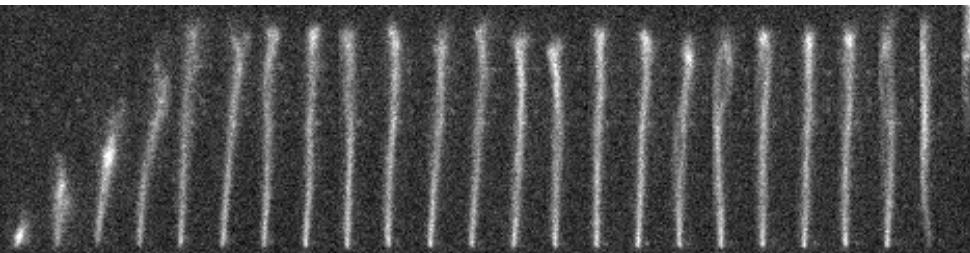


# Watching phage $\lambda$ ejection: towards a single-molecule Hershey-Chase experiment

Paul Grayson

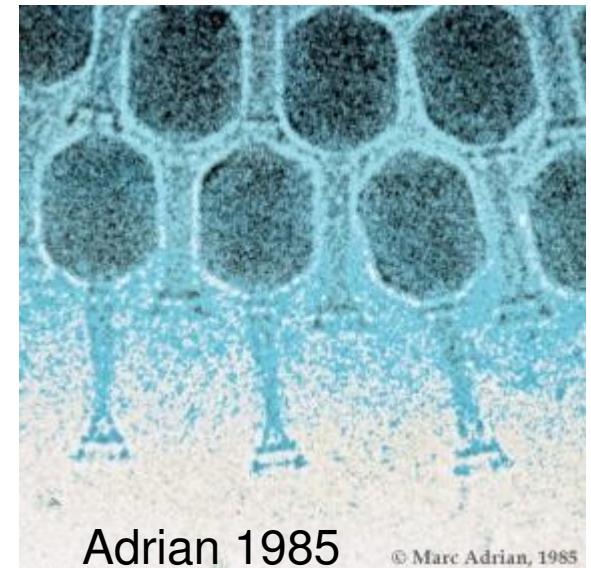
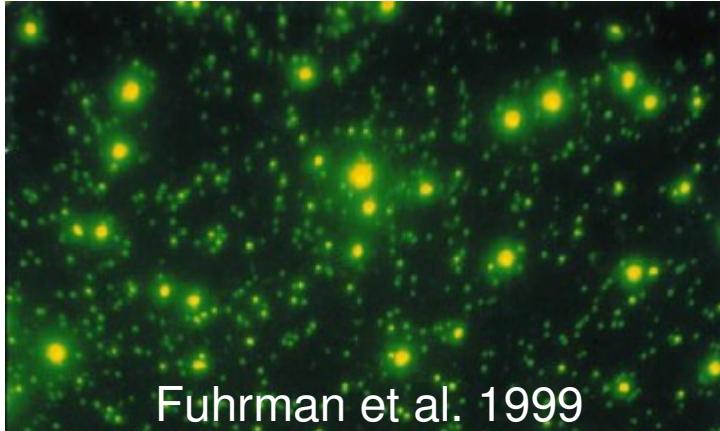
California Institute of Technology

March 14, 2007



# Bacteriophages

- Most genomes are phage genomes ( $>10^{30}$ )
- Found from polar ice to Sahara sand
- Similar to human viruses, e.g. HSV-1
- Direct causes of diseases, e.g. Cholera
- Challenge to models of evolution
- Tools for biotechnology

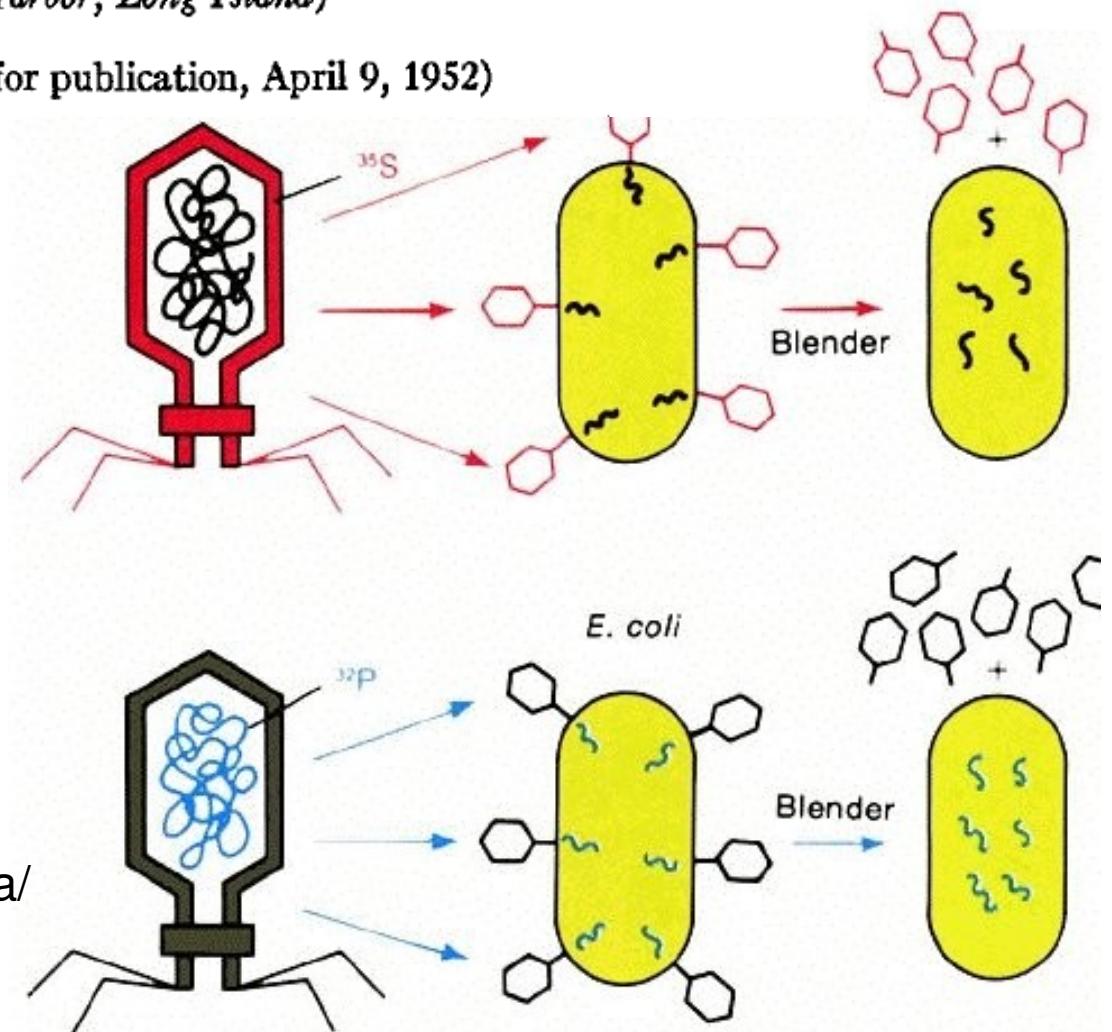


# INDEPENDENT FUNCTIONS OF VIRAL PROTEIN AND NUCLEIC ACID IN GROWTH OF BACTERIOPHAGE\*

BY A. D. HERSEY AND MARTHA CHASE

(From the Department of Genetics, Carnegie Institution of Washington, Cold Spring Harbor, Long Island)

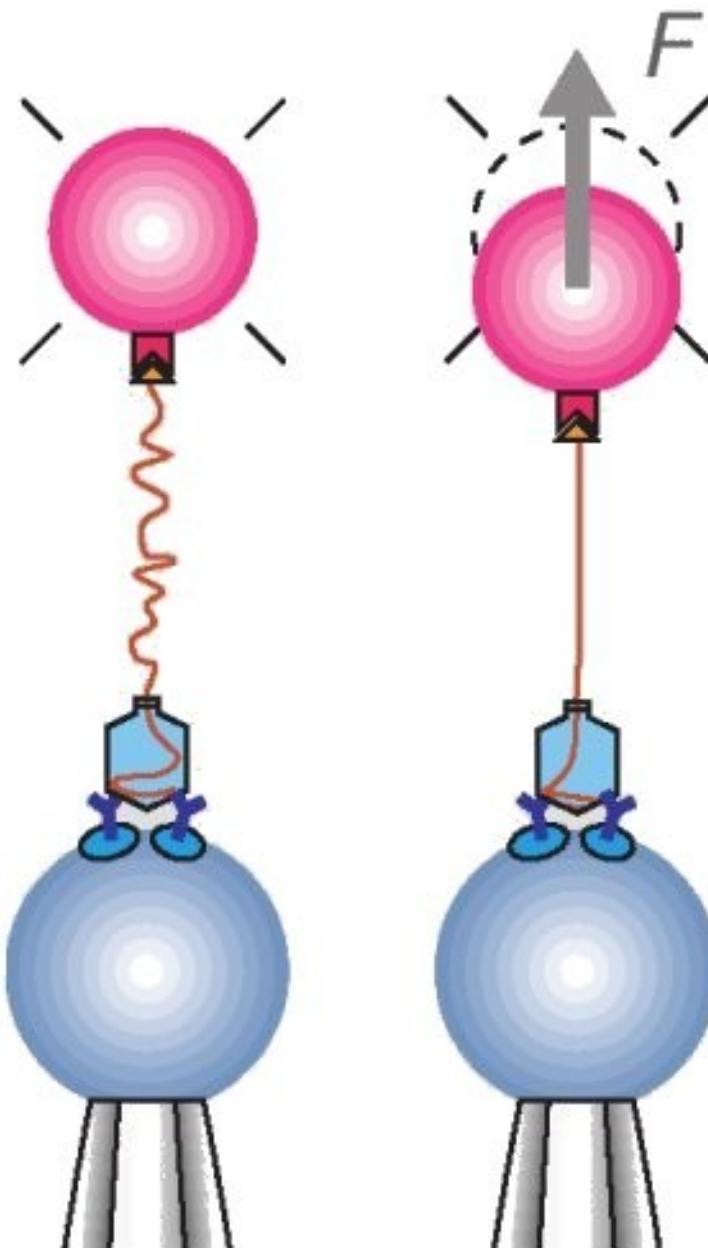
(Received for publication, April 9, 1952)



<http://www.mun.ca/>

- Hershey & Chase demonstrated DNA ejection (1952)  
...Physics asks: *how does ejection happen?*

# An inspiring experiment



Single-molecule measurements  
of phage packaging:

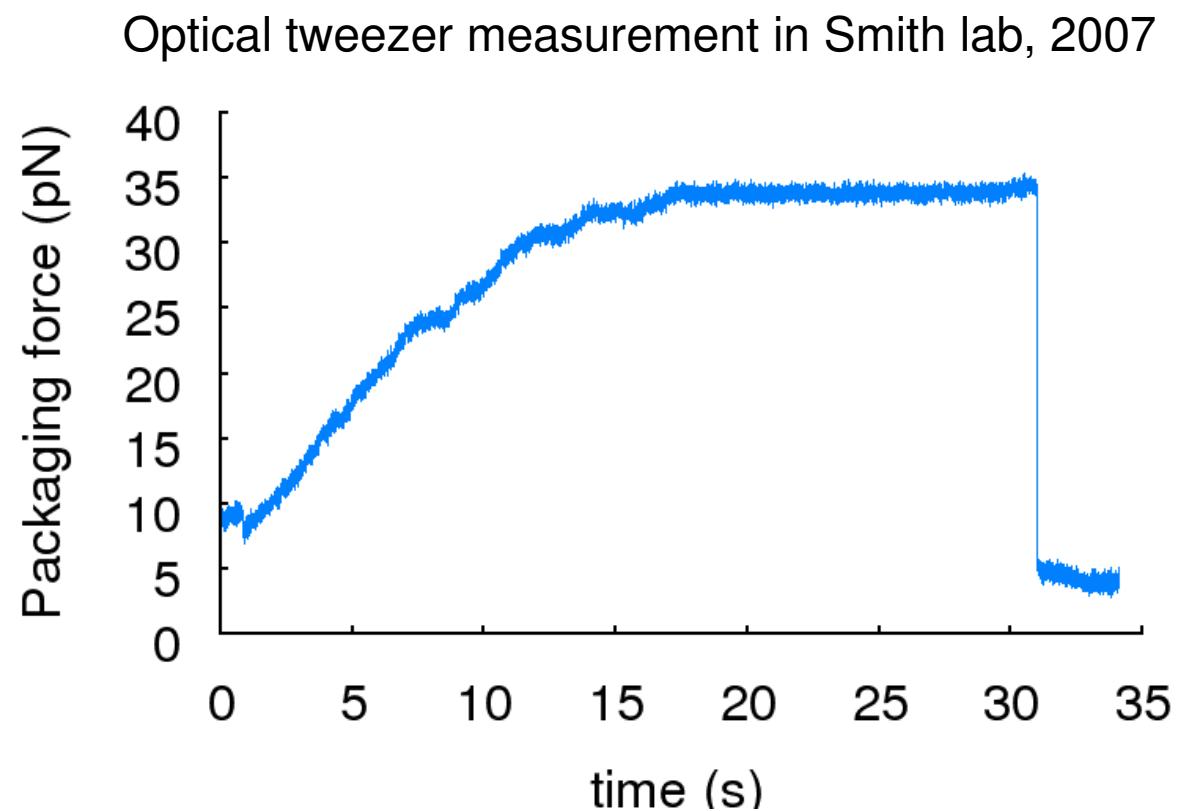
- Smith *et al.* (2001) –  $\phi$ 29 can exert  $\sim$ 60 pN  
(At 65 pN, DNA is denatured!)



# DNA packaged by a strong motor

Single-molecule measurements of phage packaging:

- 2007: The  $\lambda$  packaging motor can exert  $\sim 35$  pN !



Can we do *ejection* at the single-molecule level?

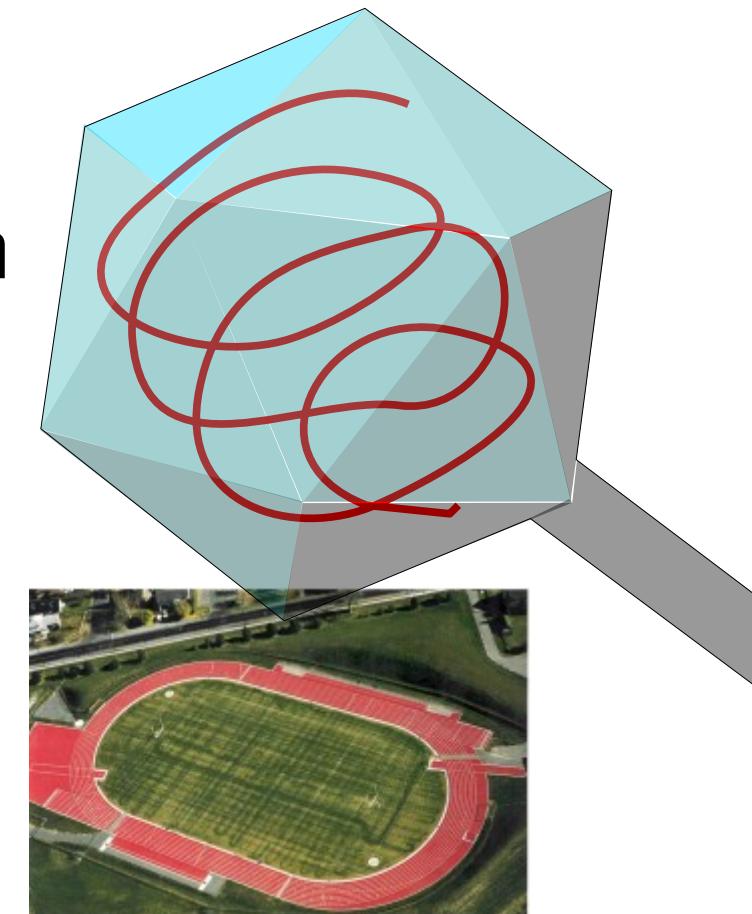
# Why so much force?

- DNA density:

16  $\mu\text{m}$  of DNA in a 57 nm capsid

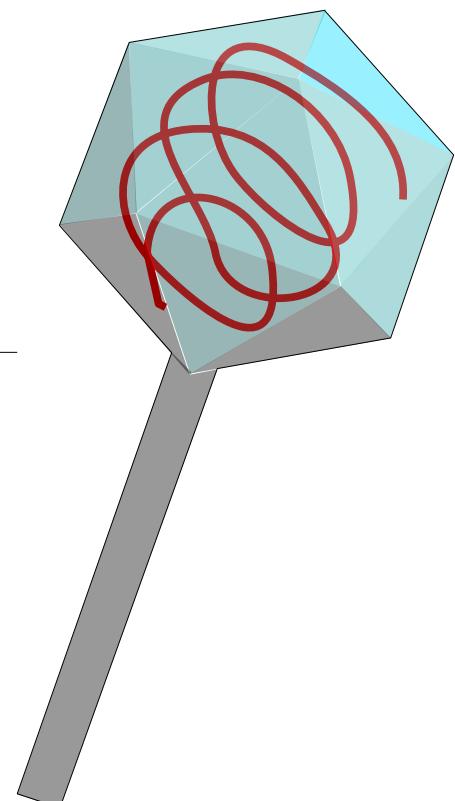
My visualization:

1 marathon in 150 m



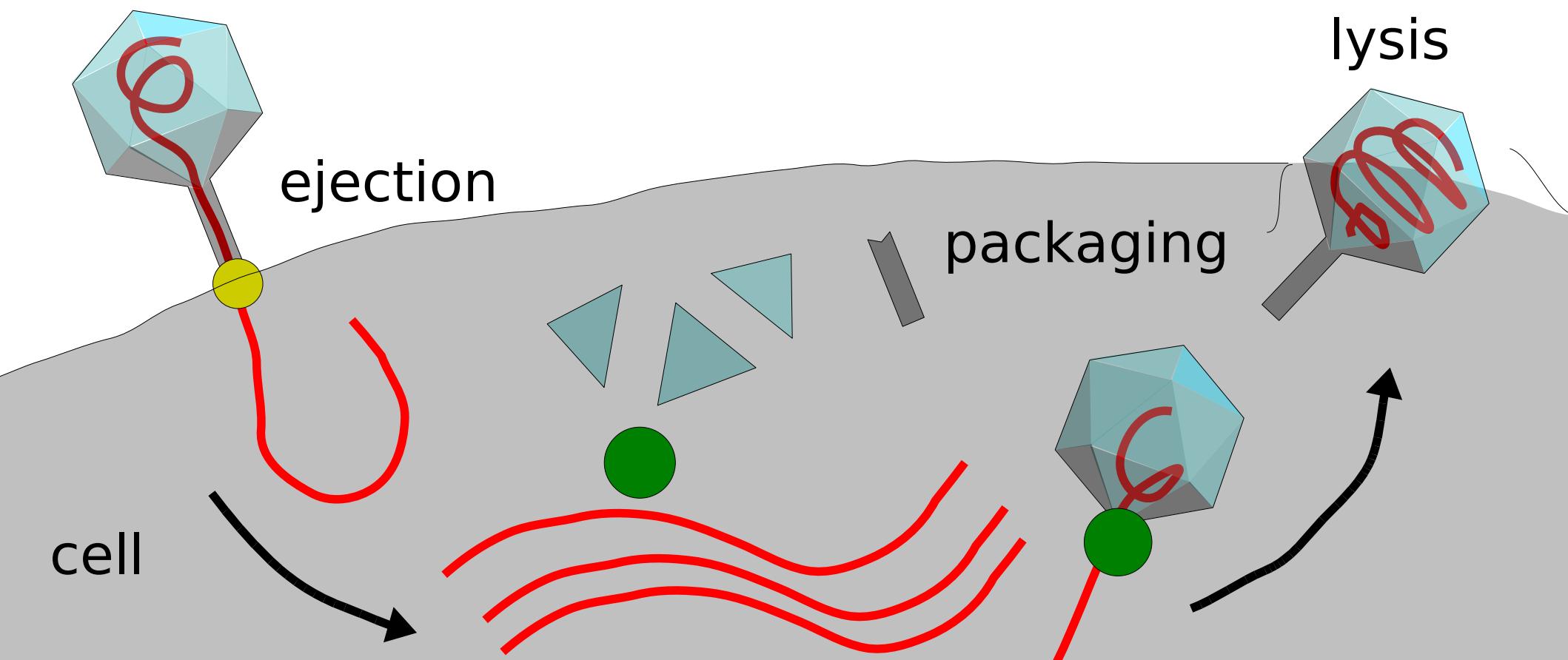
# Genome density in viruses

Virus	Genome	Capsid size	Packing density
$\lambda$	48.5 kbp	57 nm	53.4%
T7	40 kbp	55	49.0
$\phi$ 29	19.3 kbp	44.1	45.9
$\lambda$ b	38 kbp	57	41.9
polio	7.4 kb	27 nm	29.8%
HIV	18.4 kb	70	4.3
pox	186 kbp	220	3.6



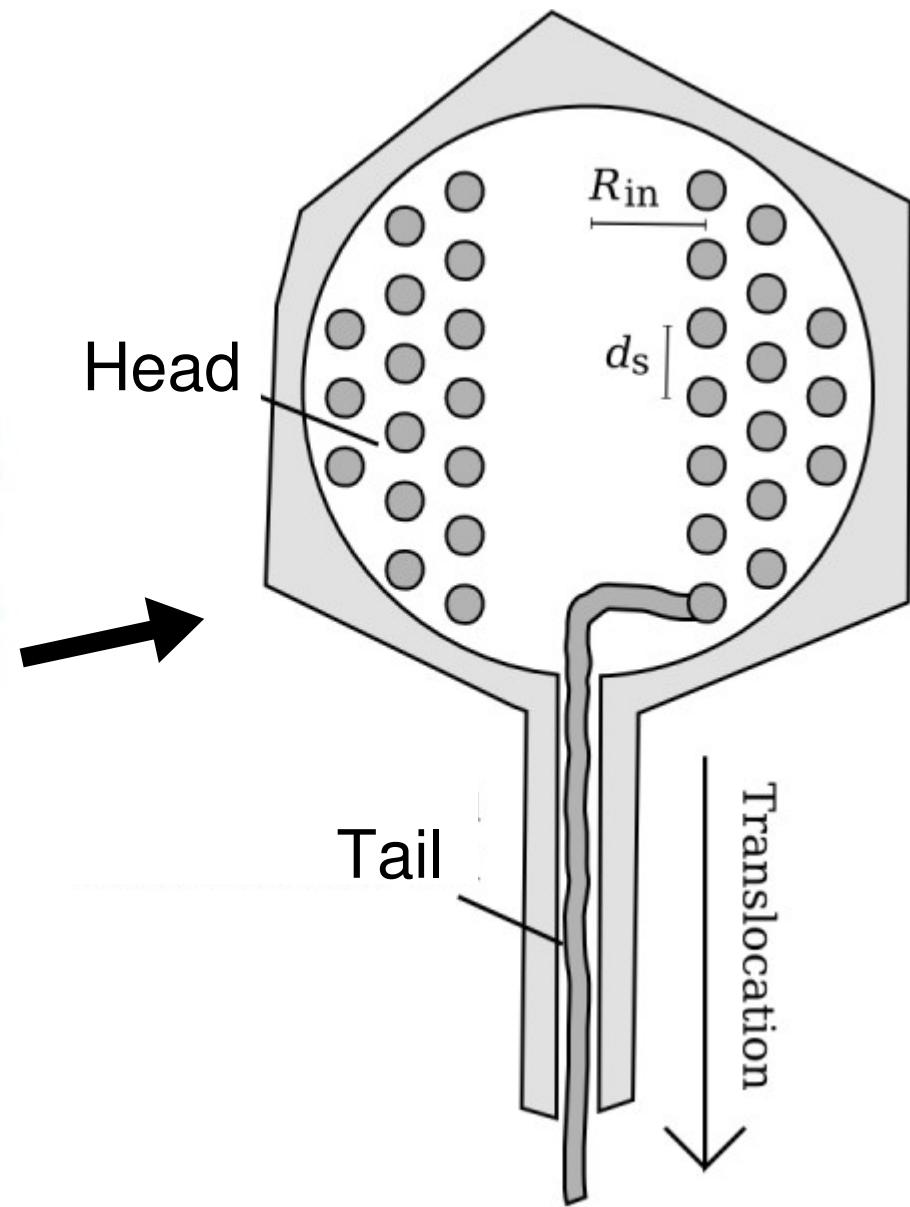
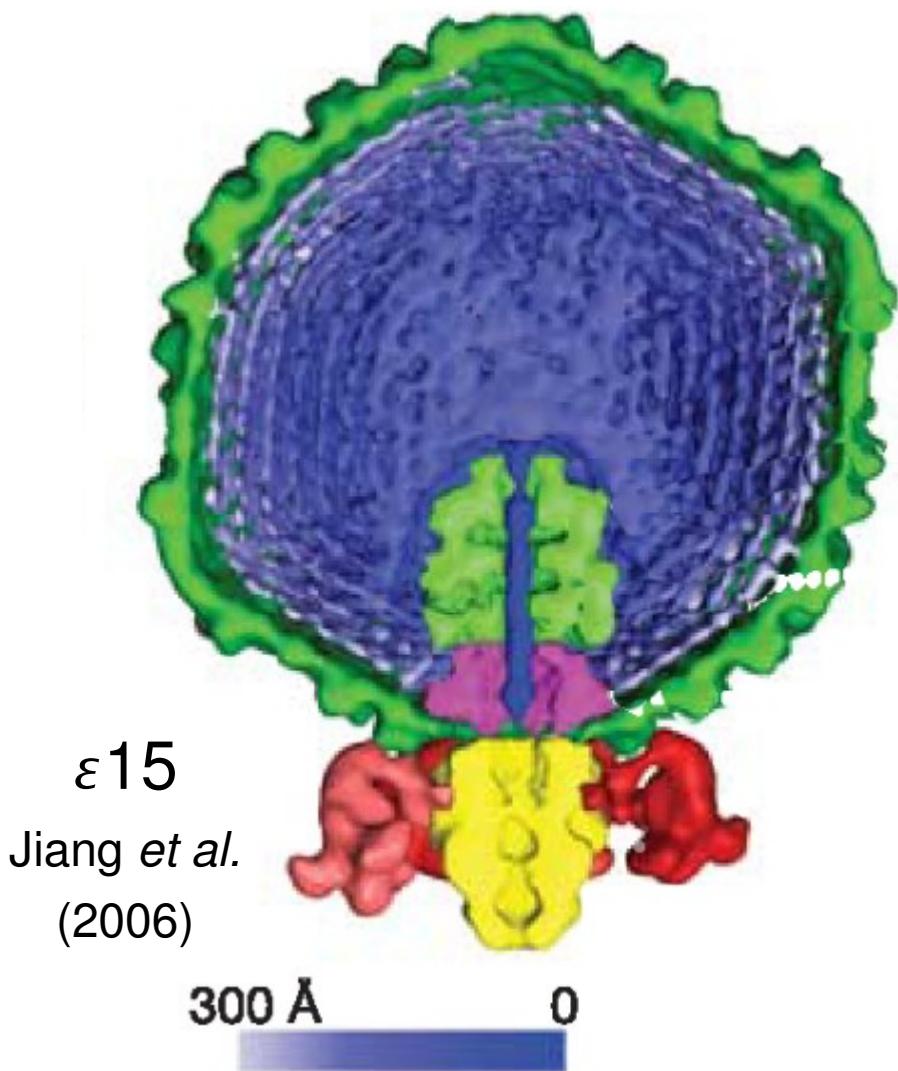
# Force is needed for ejection

- >10 pN of force, used to get DNA into the host?
- See also Grayson and Molineux, *Is phage DNA "injected" into cells - biologists and physicists can agree.* (Curr Opin Microbiology, 2007)



# Theorizing about phages

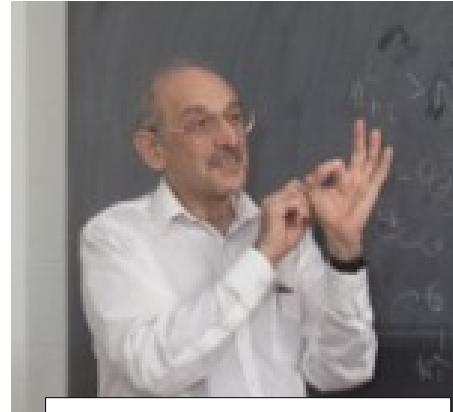
Inverse-spool model:



# Force depends on DNA density



Don Rau



Adrian Parsegian

- X-ray measurements on bulk DNA: pressure → force
- Bending stiffness relatively insignificant for the force



Theory for the ejection force with **no fitting!**

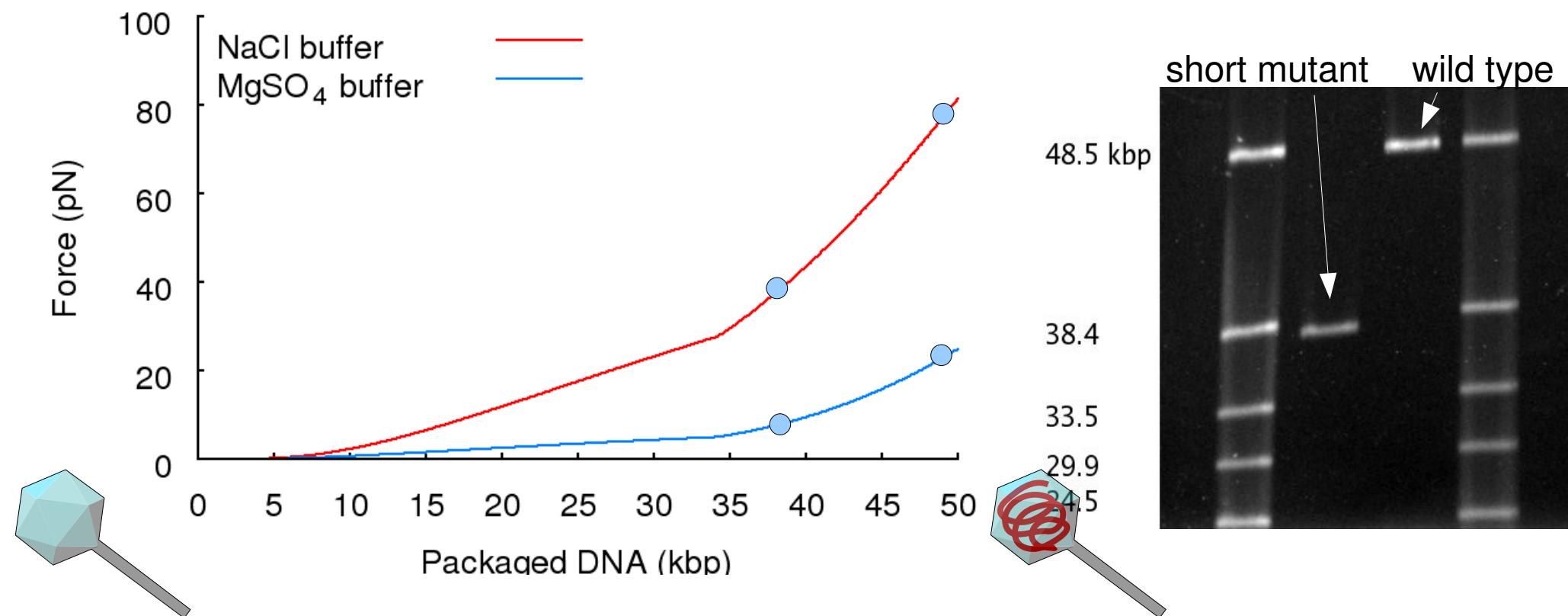
Purohit *et al.*, Forces during bacteriophage DNA packaging... (Biophys J, 2005).

Garcia *et al.*, Biological consequences of tightly bent DNA... (Biopolymers, 2007).

# Force as a function of...

Force depends on:

- Genome size (38/48 kbp)
- Ions ( $Mg^{2+}$  /  $Na^+$ )

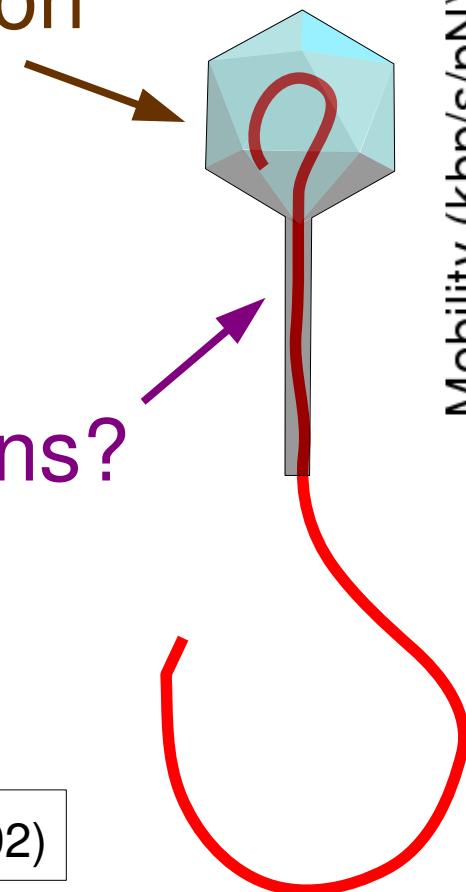


# What about dynamics?

- Define **mobility** =  $v/F$ .

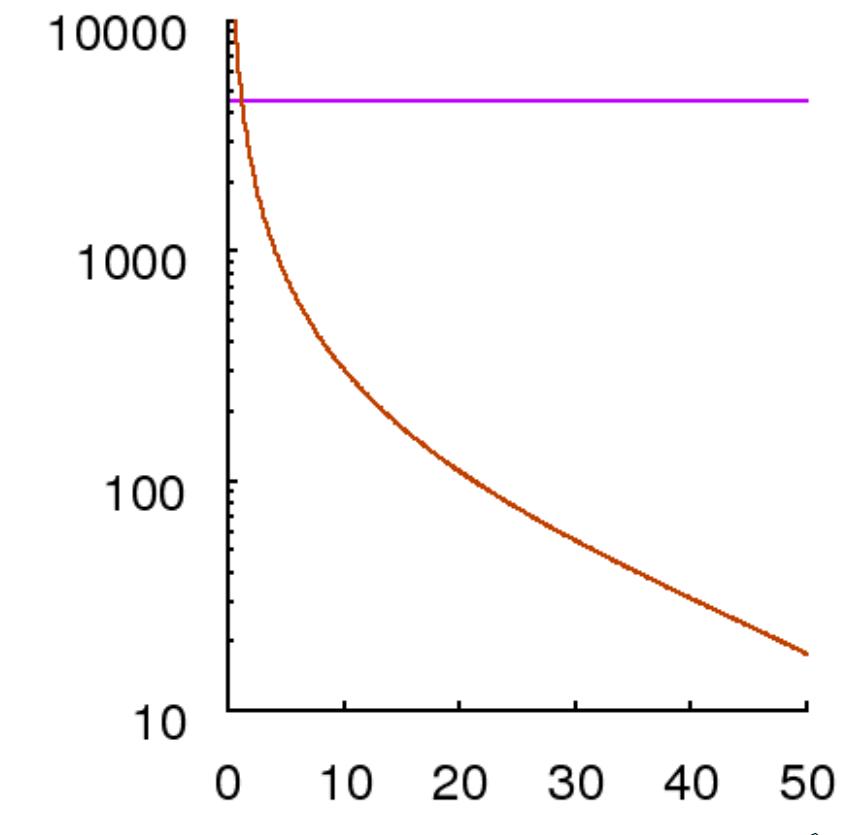
DNA-DNA interactions?

Mobility depends on  
DNA density.



DNA-tail interactions?

Constant mobility.



# Testing theory with $\lambda$ : static force

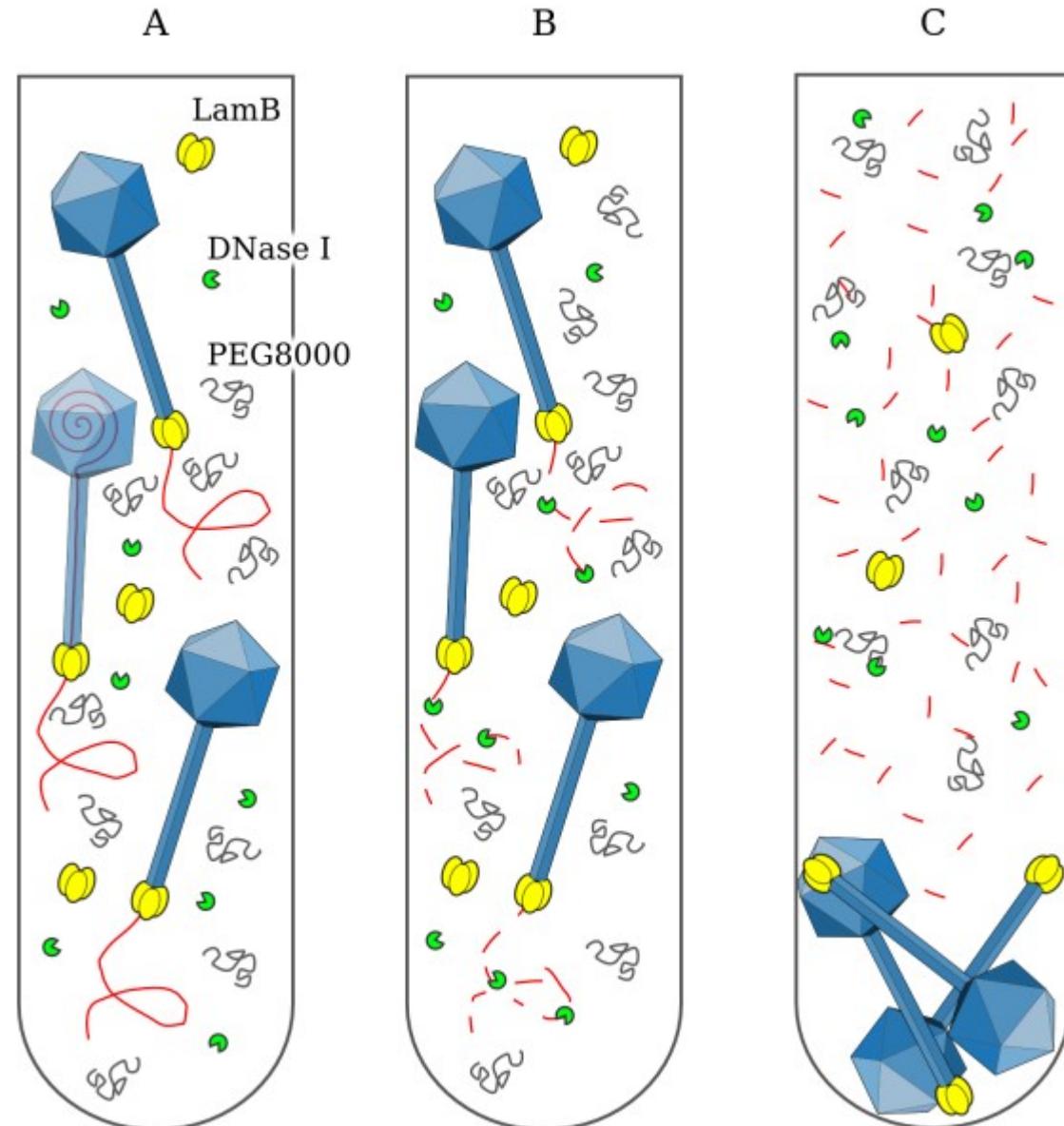
Osmotic force from  
PEG8000 balanced  
with internal force.

Bill Gelbart  
sa

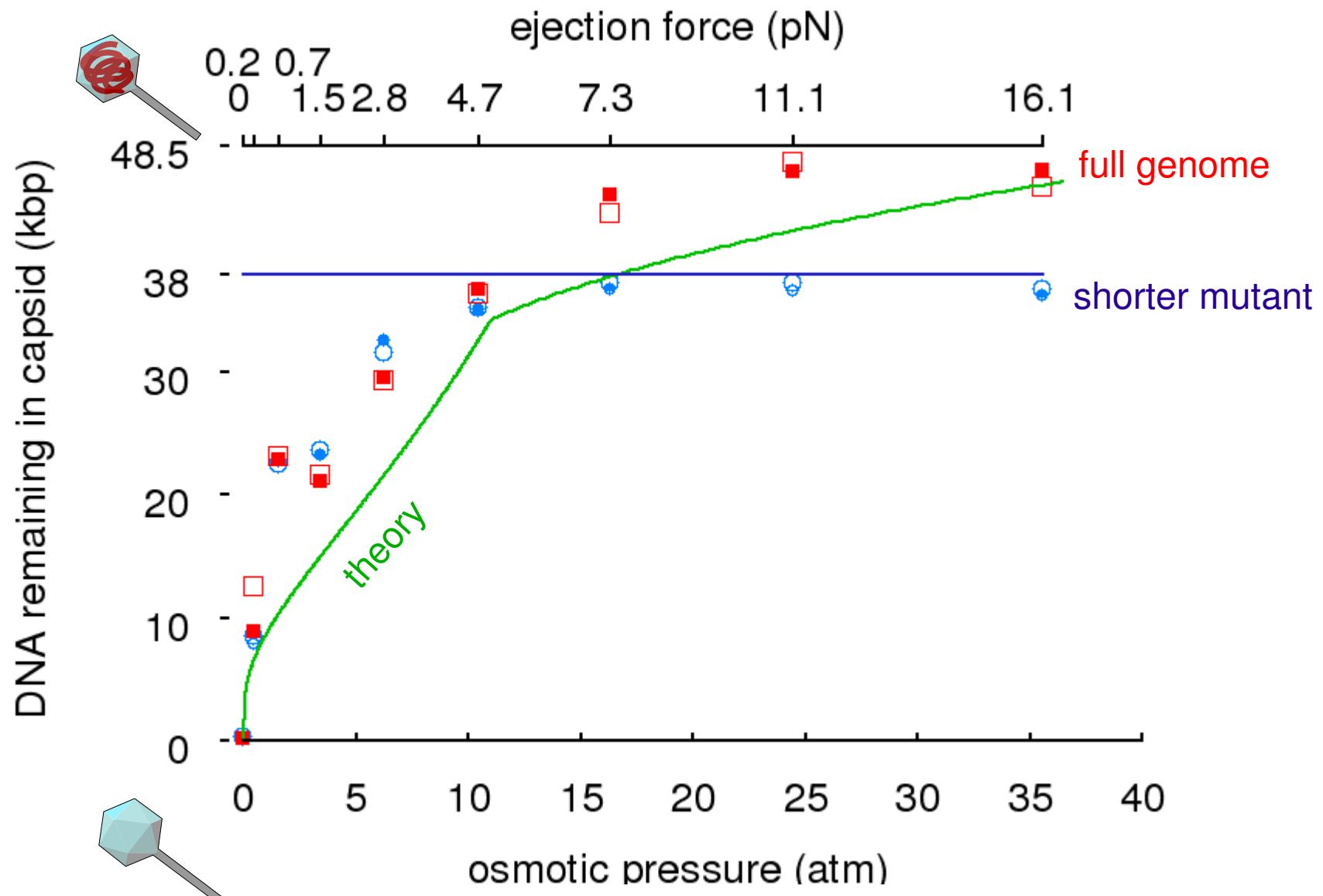


Chuck Knobler

Alex Evilevitch



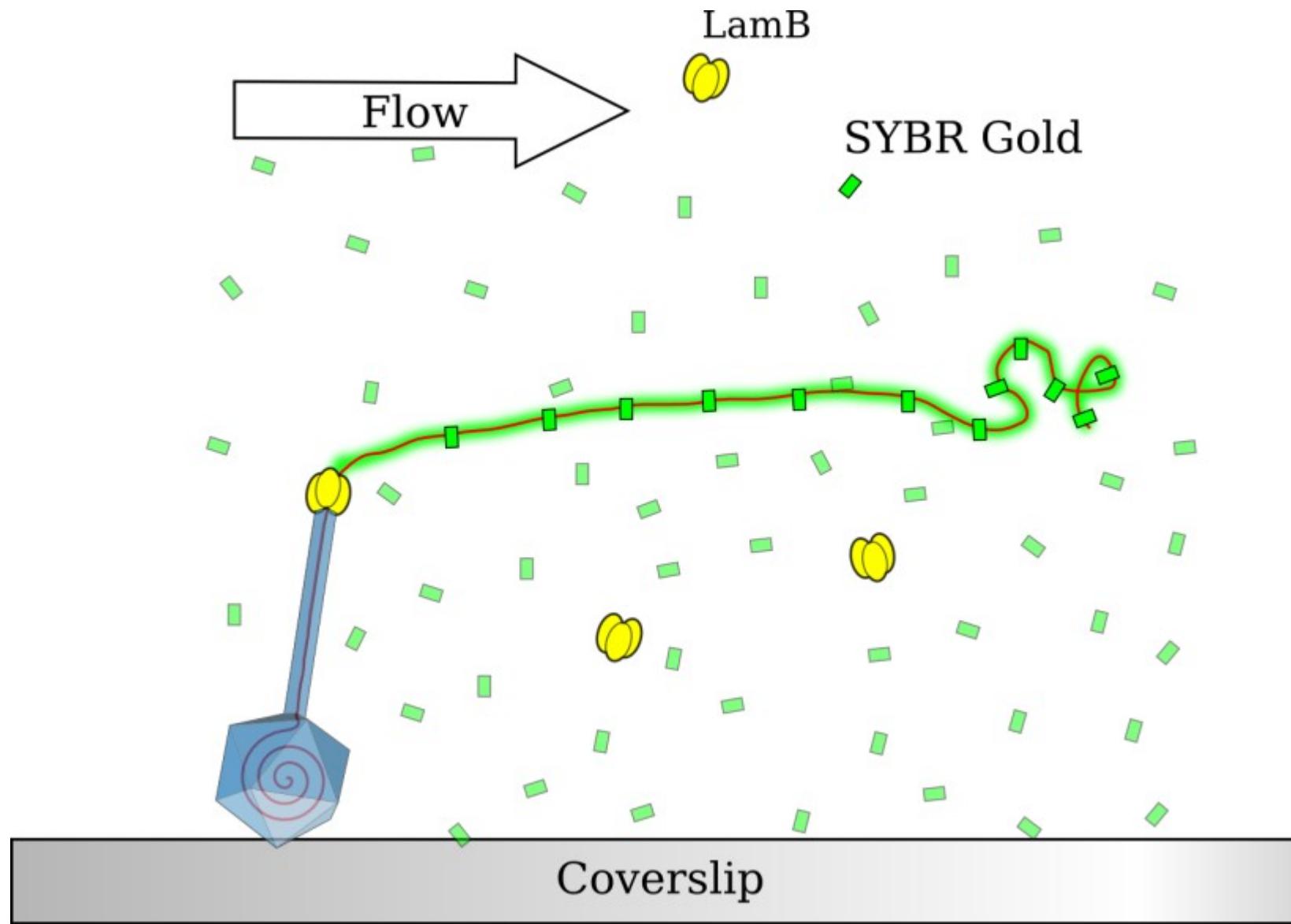
# Testing theory with $\lambda$ : static force



Grayson *et al.*, *The effect of genome length on forces in bacteriophage  $\lambda$*   
(Virology, 2006).

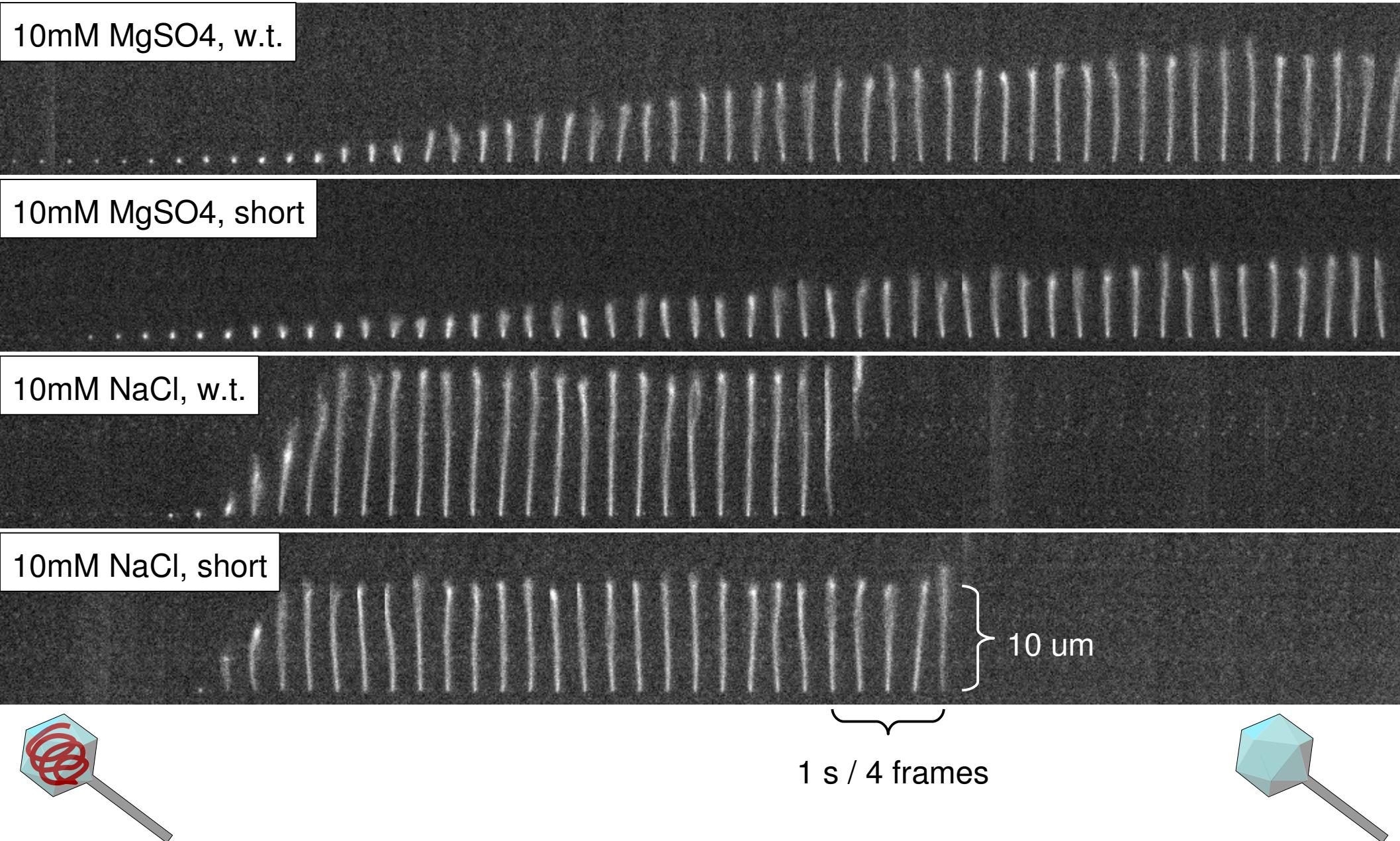
# Testing theory with $\lambda$ : dynamics

Combines methods of Mangenot *et al.* (2005) and Novick & Baldeschwieler (1988).



(show movie)

# $\lambda$ ejection trajectories



Lin Han



# Calibration with $\lambda$ fragments

10mM NaCl,  $\lambda$ /EcoRI, 3530 bp

$\lambda$ /BspHI, 7860 bp

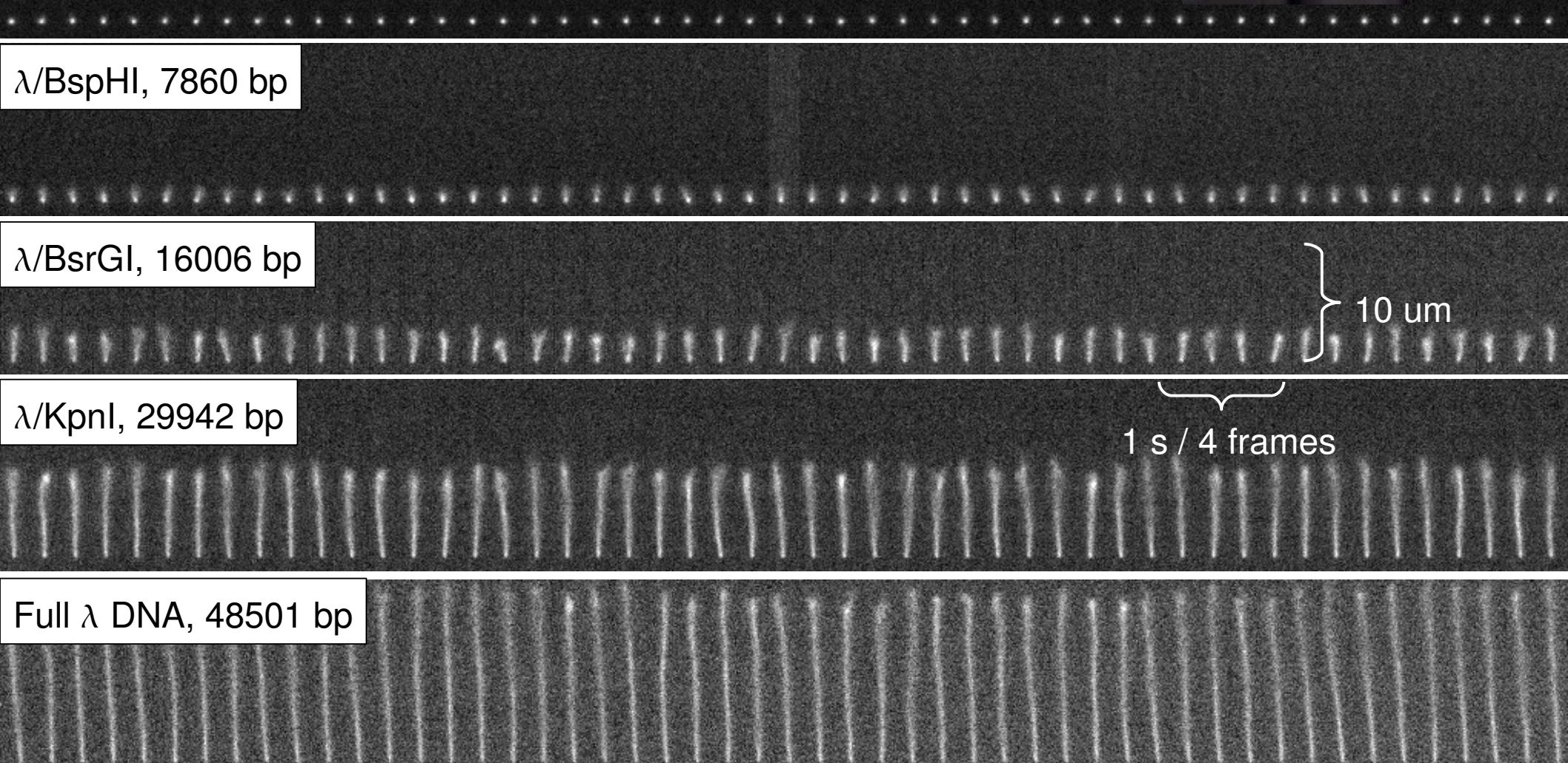
$\lambda$ /BsrGI, 16006 bp

$\lambda$ /KpnI, 29942 bp

Full  $\lambda$  DNA, 48501 bp

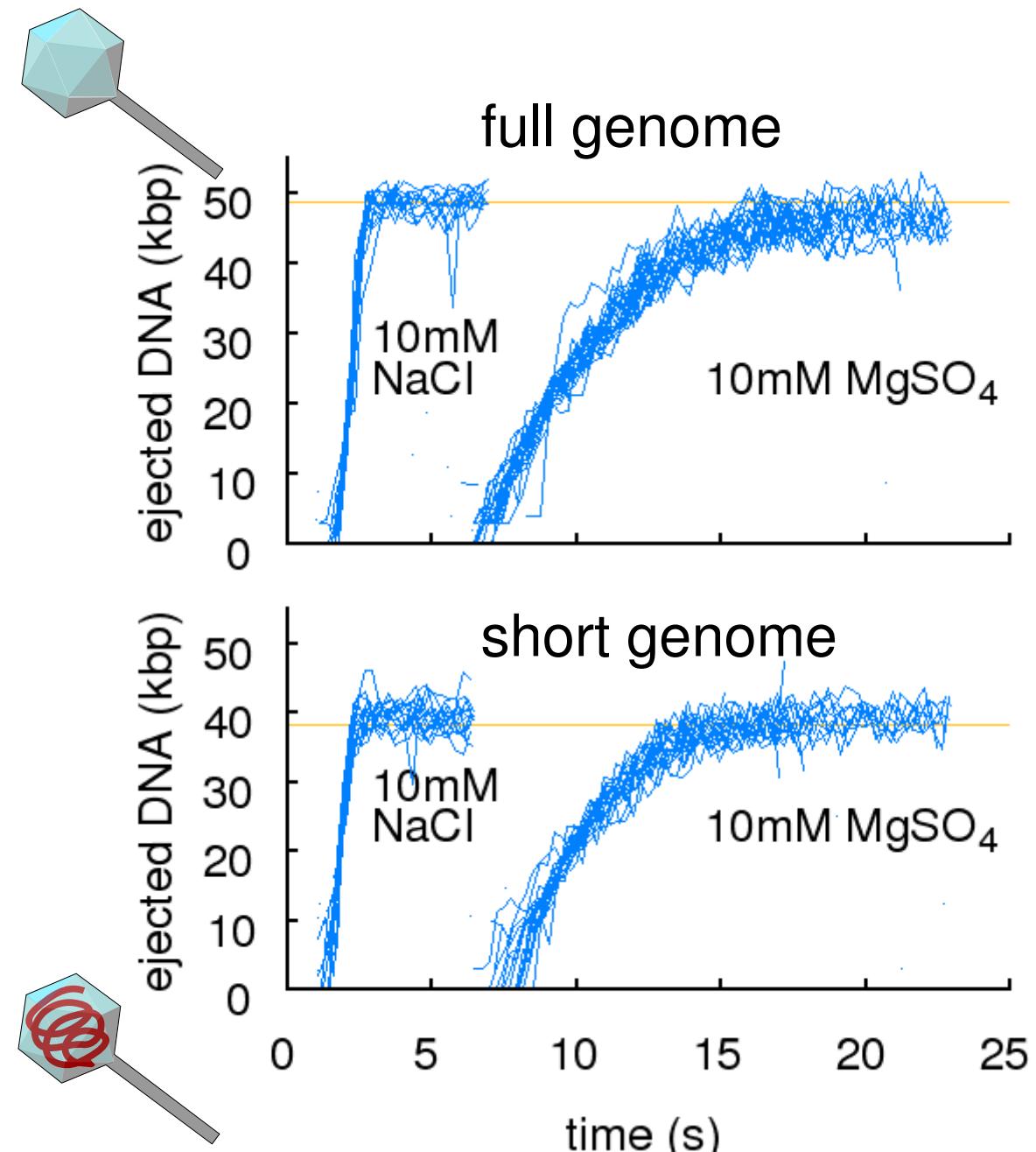
10  $\mu$ m

1 s / 4 frames

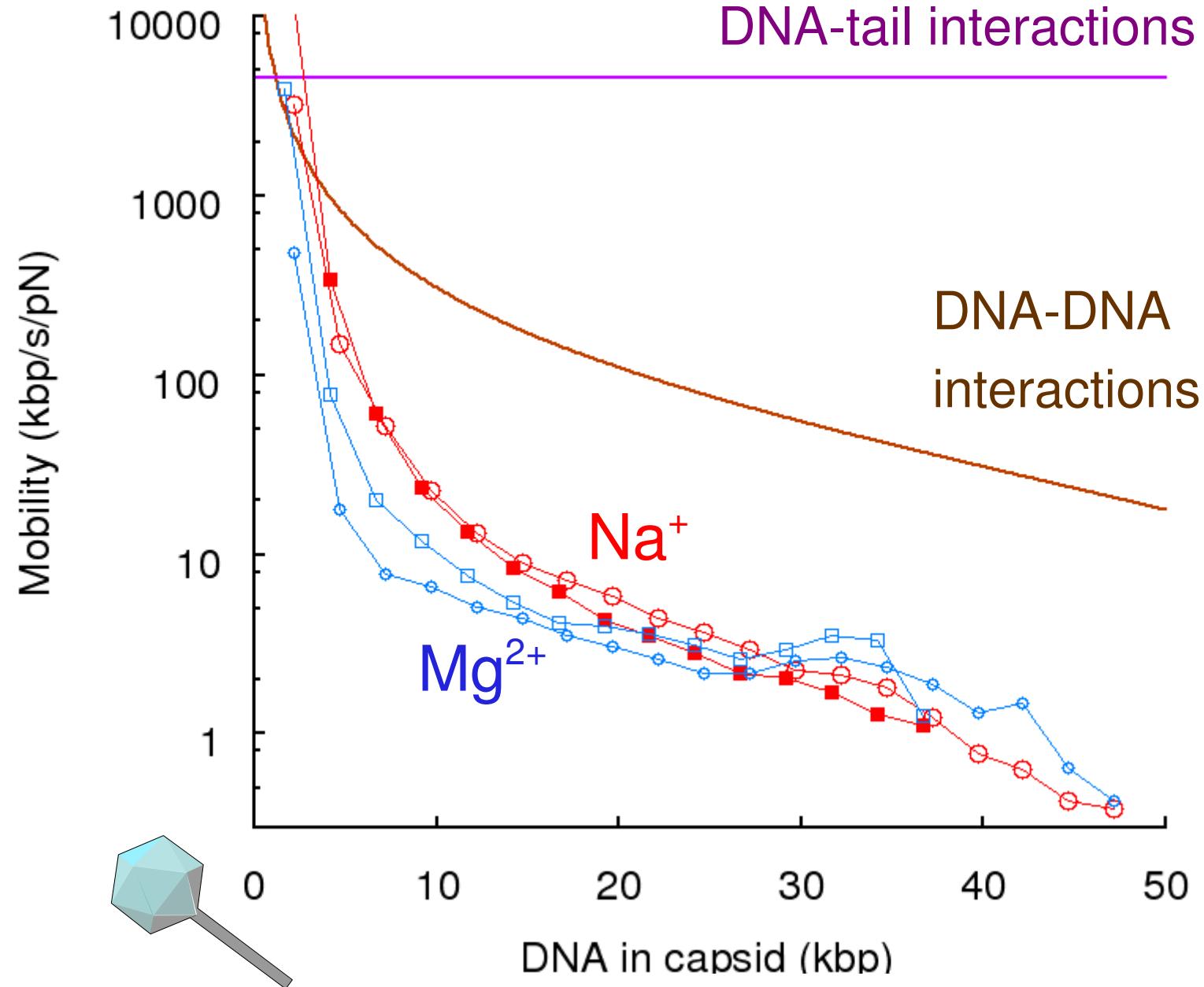


# Trajectories

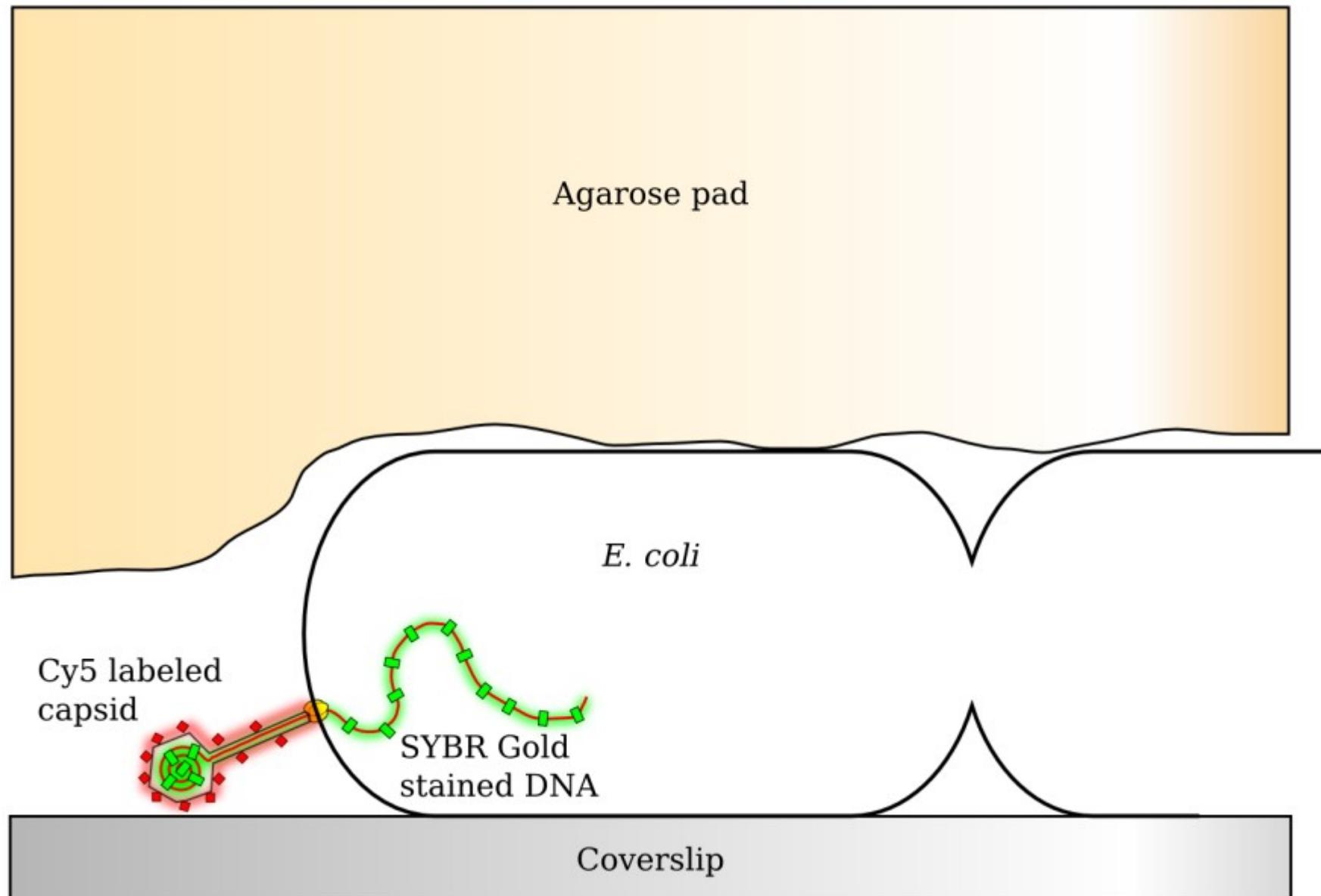
- All identical within exp. error
- Smooth motion
- Max velocity depends on ions
- Shorter phage slightly faster
- Long time at max. extension



# Mobility = $v/F$ depends on internal DNA

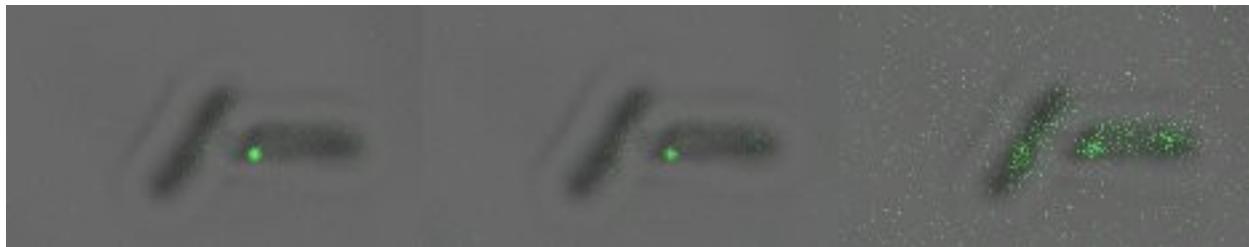


# Hershey-Chase on single phages



# Ejection *in vivo*

Label internally with SYBR Gold:

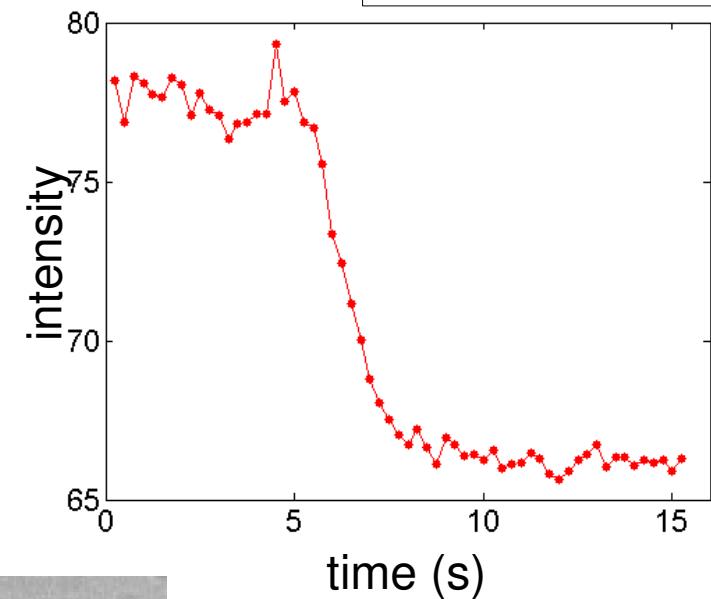
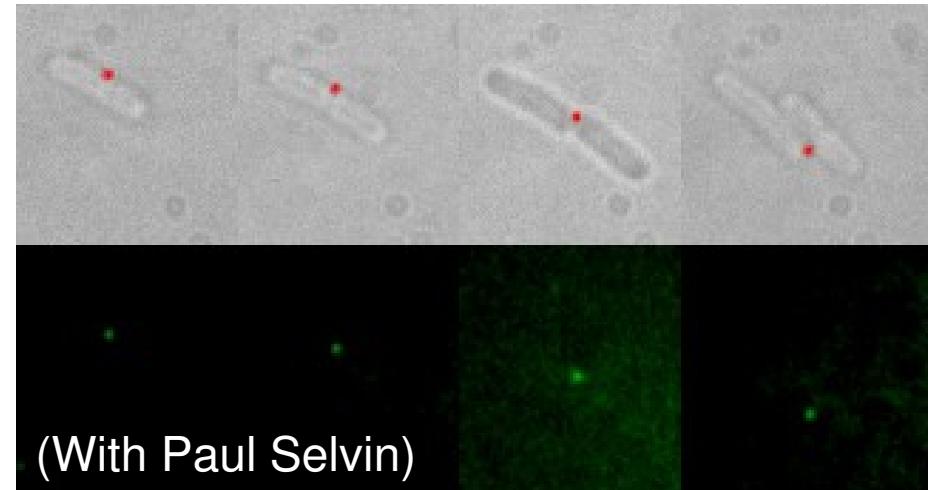


...ejection takes ~3s.

Label both DNA and protein:

Brightfield/Cy5  
(Protein)

SYBR Gold  
(DNA)



...no ejection  
in ~2h. Work  
in progress!



Tabita Winther

# Conclusions + questions

- Ejection powered by internal pressure *in vitro*.
- Friction caused by DNA-DNA interactions.
- How to quantitatively estimate friction?
- What motor completes ejection *in vivo*?

# Acknowledgments

- Collaborators: Alex Evilevitch, Lin Han, Mandar Inamdar, Bill Gelbart, Chuck Knobler, Joey Koehler, Jané Kondev, Corinne Ladous, Catie Lichten, Ian Molineux, Kelsey Nelson-James, Rob Phillips, Prashant Purohit, Erdal Toprak, Zach Travis, and Tabita Winther.s
- Fraser, Jensen, Mayo, Rees, Selvin, and Quake labs. Keck Foundation, NIH, NSF.
- Lots of help from many others!! Thanks!!!

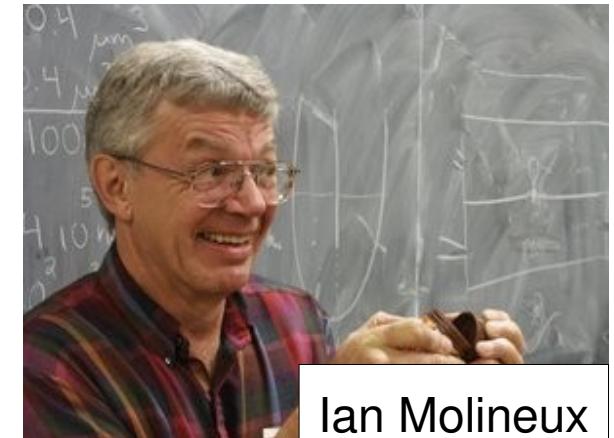
# Is pressure enough?

*E. coli* internal osmotic

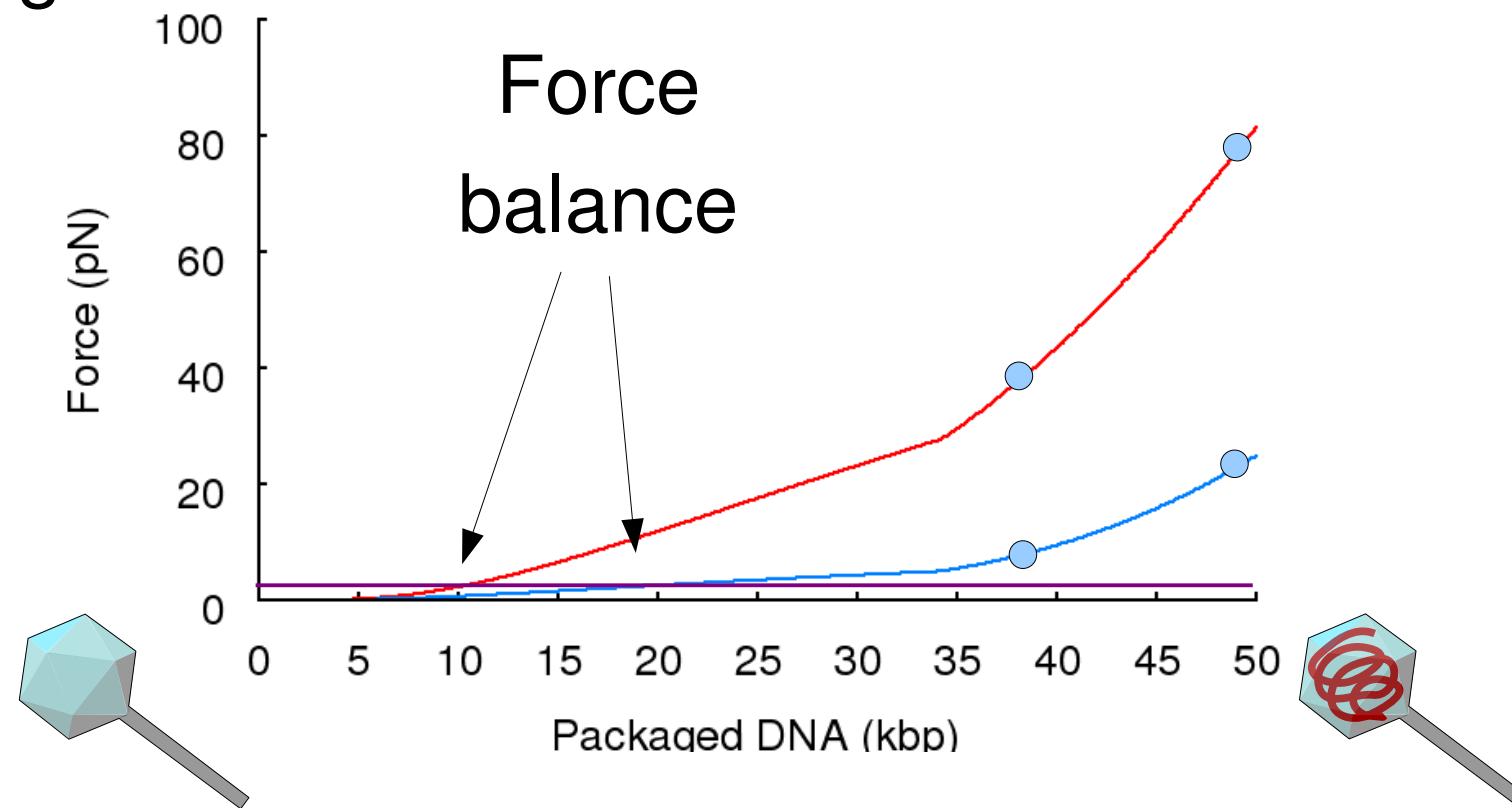
pressure = ~3 atm.

Internal motor needed!

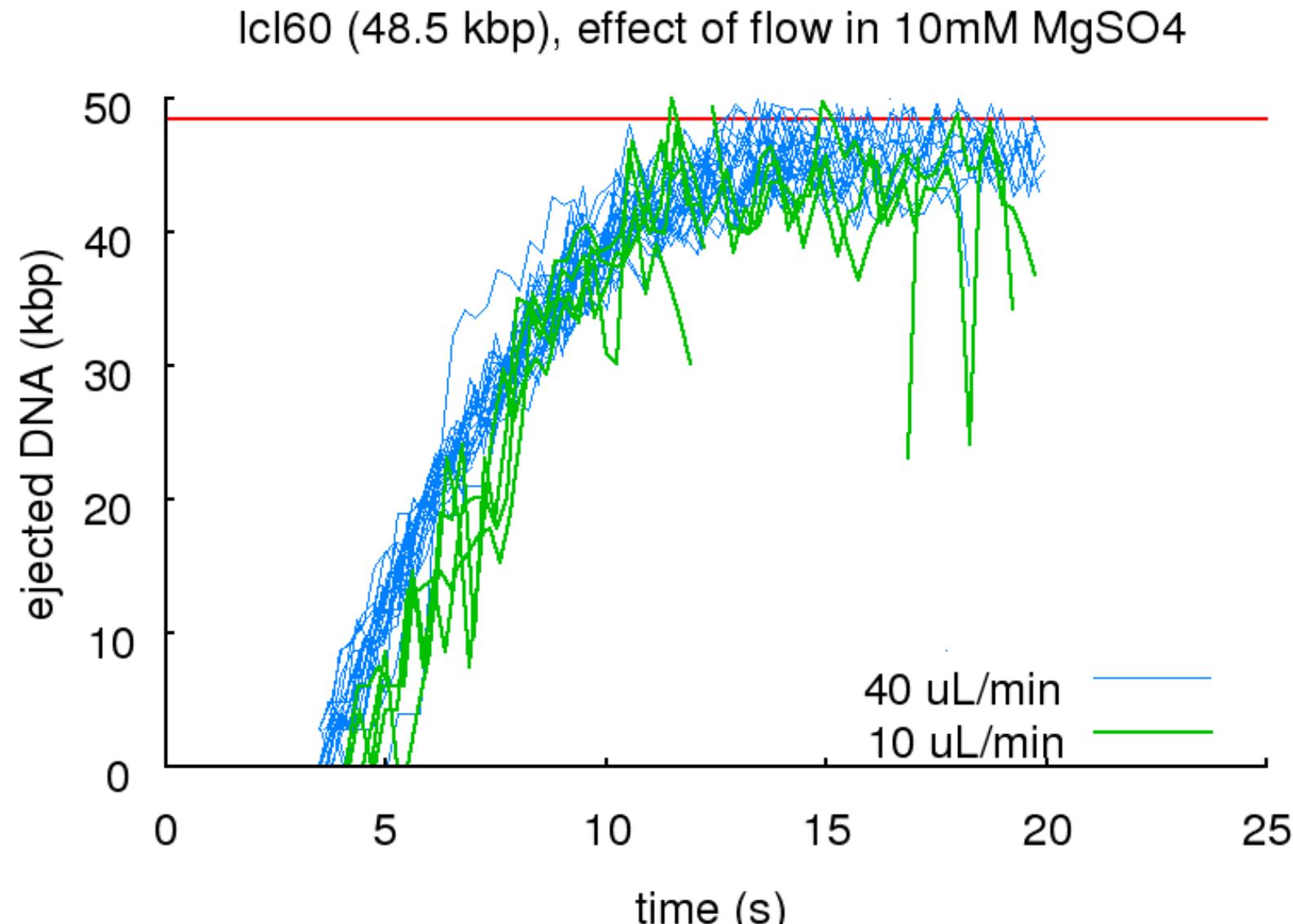
In phage T7: RNAP.



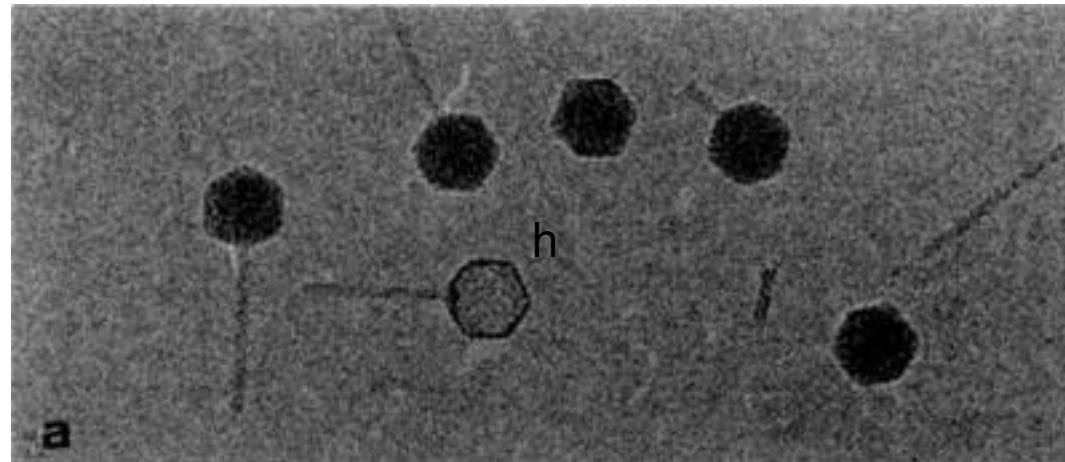
Ian Molineux



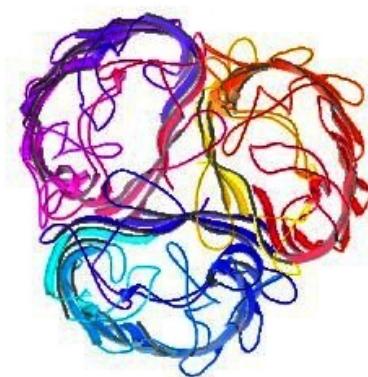
# Effect of fluid flow on ejection



# Bacteriophage $\lambda$

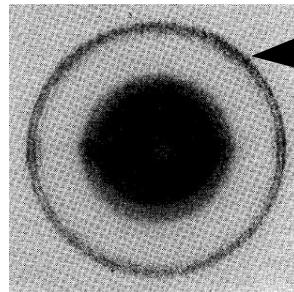


$\lambda$  capsids( Dokland & Murialdo)



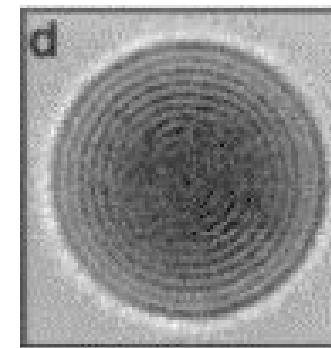
$\lambda$  receptor (LamB, maltoporin)

# Bacteriophage DNA packing

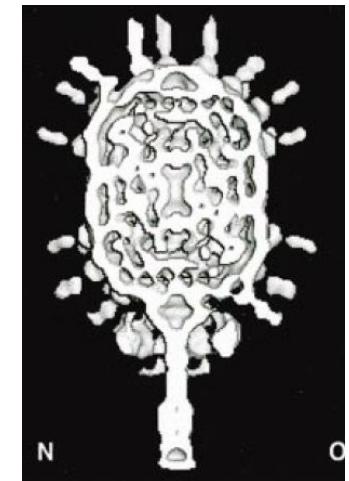


2.5 nm  
spacing

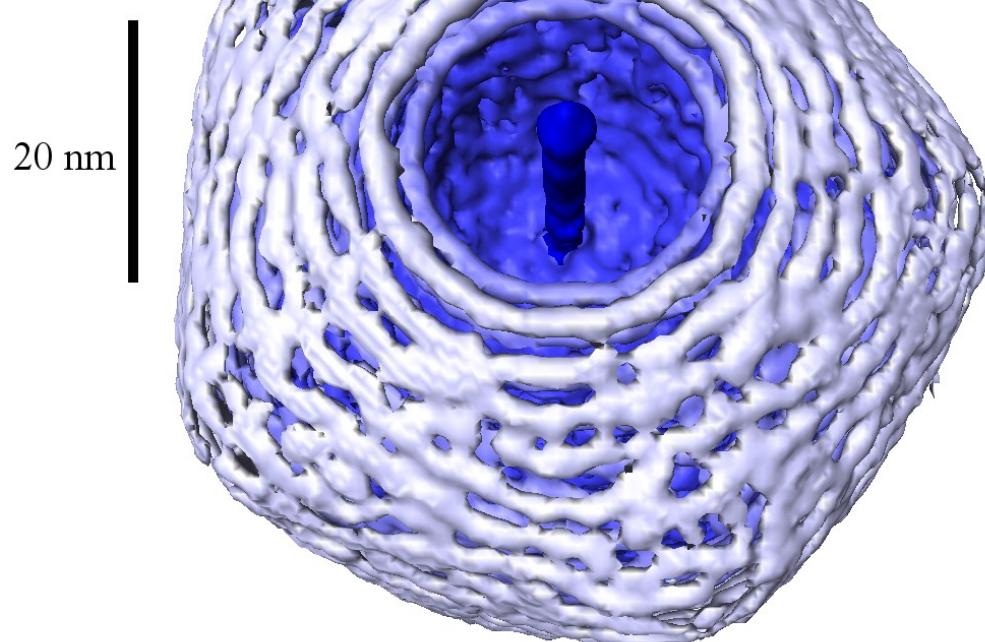
$\lambda$ , Earnshaw & Harrison (1977)  
X-ray diffraction pattern



T7, Cerritelli et al. (1997)  
averaging 10s of particles

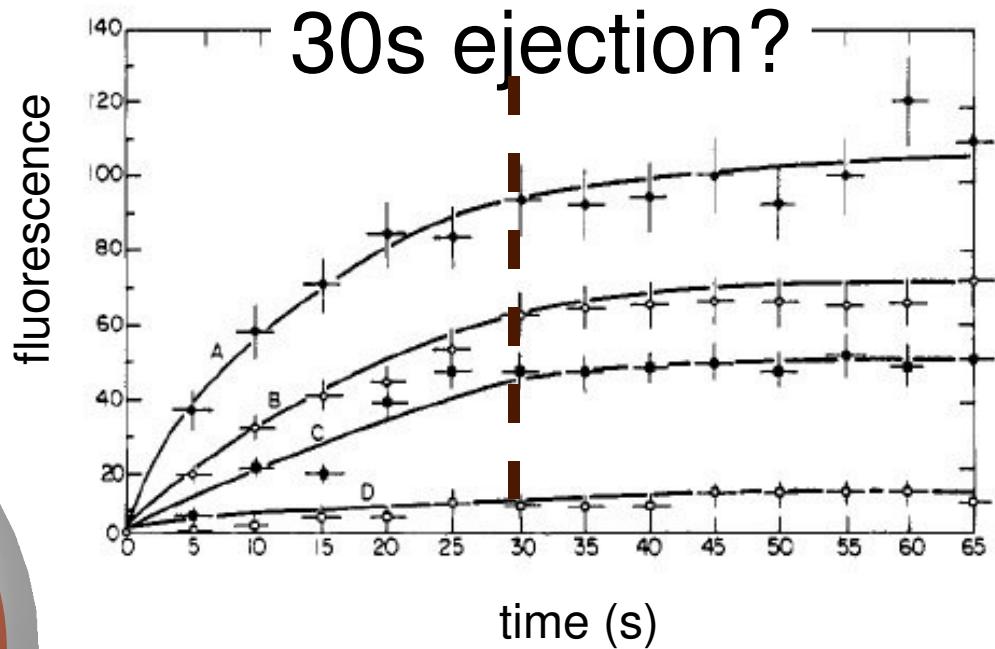
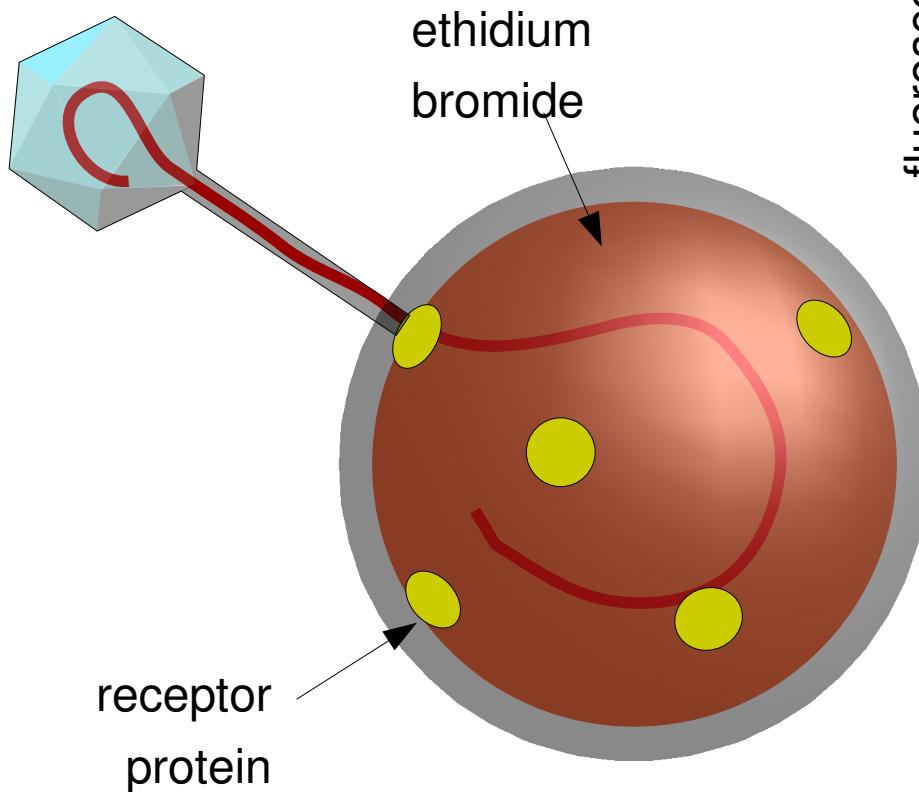


$\phi$ 29, Tao et al. (1998)  
averaging 100s of particles



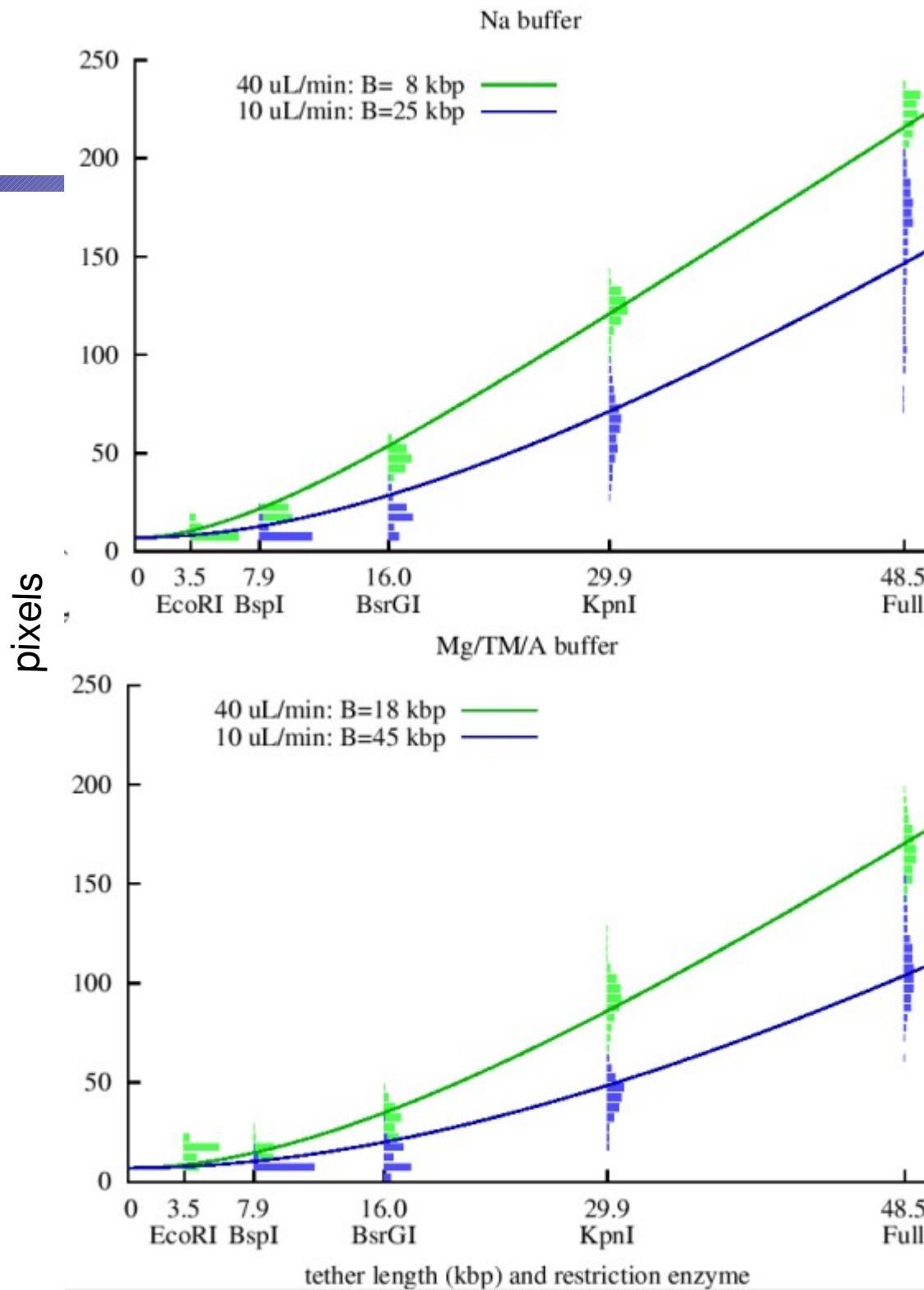
$\varepsilon$ 15, Jiang et al. (2006)  
averaging 1000s of particles

# Original rate measurements by Novick & Baldeschwieler, 1988



30s ejection?  
...but only 1000  
molecules of dye  
in each vesicle!  
This is *initiation* time.

# Calibration



# Measure velocity vs. position

